Mean velocity and effective diffusion constant for translocation of biopolymer chains across membrane

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Abstract. Chaperone-assisted translocation through a nanopore embedded in membrane holds a prominent role in the transport of biopolymers. Inspired by classical Brownian ratchet, we develop a theoretical framework characterizing such a translocation process through a master equation approach. In this framework, the polymer chain, provided with reversible binding of chaperones, undergoes forward/backward diffusion, which is rectified by chaperones. We drop the assumption of timescale separation and keep the length of a polymer chain finite, both of which happen to be key points in most of the previous studies. Our framework makes it accessible to derive analytical expressions for mean translocation velocity and an effective diffusion constant in a stationary state, which is the basis of a comprehensive understanding of the dynamics of such a process. Generally, the translocation of polymer chains across a membrane consist of three subprocesses: initiation, termination, and translocation of the main body part of a polymer chain, where the translocation of the main body part depends on both the binding/unbinding kinetics of chaperones and the diffusion of the biopolymer chain. This is the main concern of this study. Our results show that the increase of the forward/backward diffusion rate of a polymer chain and the binding/unbinding ratio of chaperones both raise the mean translocation velocity of a polymer chain, and the mean velocity finally reaches a saturation amount with an extremely rapid diffusion or extremely high binding/unbinding ratio. Roughly speaking, the dependence of effective
Translocation across membranes are ubiquitous during or after the synthesis of biopolymers. A typical example is of proteins that translocate across the endoplasmic reticulum through an aqueous-pore protein-conducting channel [1–5]. Additional examples, including the exportation of RNA across nuclear membranes in eukaryotic cells [6, 7] and the viral injection of DNA into a host [8, 9], both involve translocation through nanoscopic pores embedded in biological membranes. New evidence also indicates that the uptake of long DNA molecules concern the linear passage through the nuclear pore complex [10]. Despite the lack of a membrane-bound nucleus or any membrane-bound organelle, the DNA of bacteria travels across the cell envelope through narrow constrictions during the process of horizontal gene transfer [11–13], such as transformation [14], which is responsible for the adaptive evolution of bacteria. Similar transport mechanisms have also been reported in the biotechnology of drug delivery [15] and rapid DNA sequencing [16].

In recent decades, wide attention has been devoted to understand the mechanism of such crucial biological processes. Pioneering work has established various mechanistic understandings of what induces the directional movement of biopolymers through narrow pores. One important driving force is the external electric field across the membrane, demonstrated by in vitro experiments [17, 18] and extensive theoretical analysis.

Keywords: biomolecules, molecular motors, systems biology

References

1. Introduction

Translocation across membranes are ubiquitous during or after the synthesis of biopolymers. A typical example is of proteins that translocate across the endoplasmic reticulum through an aqueous-pore protein-conducting channel [1–5]. Additional examples, including the exportation of RNA across nuclear membranes in eukaryotic cells [6, 7] and the viral injection of DNA into a host [8, 9], both involve translocation through nanoscopic pores embedded in biological membranes. New evidence also indicates that the uptake of long DNA molecules concern the linear passage through the nuclear pore complex [10]. Despite the lack of a membrane-bound nucleus or any membrane-bound organelle, the DNA of bacteria travels across the cell envelope through narrow constrictions during the process of horizontal gene transfer [11–13], such as transformation [14], which is responsible for the adaptive evolution of bacteria. Similar transport mechanisms have also been reported in the biotechnology of drug delivery [15] and rapid DNA sequencing [16].

In recent decades, wide attention has been devoted to understand the mechanism of such crucial biological processes. Pioneering work has established various mechanistic understandings of what induces the directional movement of biopolymers through narrow pores. One important driving force is the external electric field across the membrane, demonstrated by in vitro experiments [17, 18] and extensive theoretical analysis.
Mean velocity and effective diffusion constant for translocation of biopolymer chains across membrane

[19–21]. Considering there is usually not a strong enough electric field in living cells, the other mechanism, Brownian ratchet suggests pure random thermal diffusion is rectified by chemical asymmetry between the cis and trans sides of the membrane, giving rise to directional motion [22, 23]. This is the main concern of this paper. A typical case, particularly for proteins, is that the binding of chaperones to the translocating polymer on the trans side of a membrane prohibits the polymer chain’s backward diffusion to the cis side, thus leading to the directional translocation [3, 24, 25]. This mechanism appears to be confirmed on the basis of empirical data for the transport of prepro-α-factor into the lumen of endoplasmatic reticulum, where prepro-α-factor is regarded as the protein and BiP serves as a chaperone molecule [26]. Recent progress on the kinetic uptake of DNA also provides evidence for analogous mechanisms [27].

Quantitative investigations devoted to chaperone-assisted transcriptions mainly focus on the dynamic properties of the system, with theoretical explorations mostly setup through a continuous space and time description [22, 23, 28]. Such continuum models can work effectively in the limit that chaperones are much larger compared with the translocating polymer. However, this may become unrealistic in living cells.

Other studies contribute to translocation process of such structure usually through discrete master equations. Early results based on one-dimensional master equations are derived on the simple assumption that both the attachment and the detachment of corresponding chaperones are much faster than the translocation rate, leaving the state of individual sites drawn from the stationary distribution [29, 30]. Dropping this approximation, a more general discrete model is then proposed [31], where the translocating polymer is represented by a one-dimensional lattice of infinite length and an analytical expression for the transcription velocity is obtained mathematically. Extensive analysis was carried out on this model to determine the diffusion constant and even higher cumulants of the translocated length [32], while the same process is discussed from the viewpoint of first-passage time [33]. These three models enjoy a common feature, they always keep track of the state of each binding site that has been translocated to the target region. This approach, although ideal, does not seems feasible, especially for long polymer chains, since the space of states shall experience an explosion as the number of binding sites grows. In this sense, such models makes a difference merely on the numerical simulation side, even if explicit expressions for some dynamic quantities can be obtained. To avoid this issue, a fresh study on this problem setup a modified translocation ratchet model involving the state of only a few binding sites [34]. Although a model is proposed dropping the key assumption of timescale separation between the binding/unbinding process of chaperones and the forward/backward diffusion process, analytical results become no longer accessible. Meanwhile, the length of a particular polymer chain is always assumed to be infinite on both sides of the membrane in this model, which is not quite appropriate.

In this paper, based on the classical Brownian ratchet, a general framework describing the chaperone-assisted translocation of a polymer chain across a membrane is provided. Since the binding/unbinding kinetics of chaperones might have a sizable effect on the dynamics of the translocation process, as addressed in previous work [31, 32], the state of binding sites located just behind the membrane, i.e. whether it is bound with a chaperone, is taken into account explicitly in our model. Moreover, the usual assumption of infinite length of polymer chain is also rejected. That is to say,
Mean velocity and effective diffusion constant for translocation of biopolymer chains across membrane

the number of binding sites of a particular polymer chain, or the length of a polymer chain, is assumed to be finite in our model. Through a scheme similar to that in previous investigations [35], the mean velocity and effective diffusion constant of polymer translocation are obtained explicitly.

This paper is organized as follows. On the concept of Brownian ratchet, a model describing the process of polymer translocation across membrane is introduced in section 2. How this model works will be explained in detail with coupled master equations. In section 3, we deduce analytical expressions for mean translocation velocity and effective diffusion constant. To show basic properties of the translocation process, numerical results are presented in section 4. Finally, we summarize our results briefly in section 5.

2. Theoretical description of polymer translocation across a membrane through a nanopore

During the translocation across a membrane, a polymer chain moves stochastically to the other side of the pore embedded in the membrane, which is the only possible way that polymers can pass through. Chaperones are assumed to exist in the target region only. Once the binding site of a polymer chain adjacent to the pore is occupied, the polymer chain is prevented from moving backward through the pore due to the large relative size of the chaperone with respect to the diameter of the narrow pore.

A polymer chain of finite length is represented as a one-dimensional lattice with $N + 1$ binding sites, labeled $0, 1, 2, \cdots, N$. For convenience, the distance between each two neighboring binding sites is normalized to one. If we take the polymer chain as a reference frame, the translocation process of a polymer chain across membrane through a pore can be regarded as the membrane’s stochastic motion along a one-dimensional lattice. We represent the membrane as a wall for simplicity. If the wall is standing at site $k$ and the previous site, labeled $k - 1$, remains unoccupied, the wall may hop stochastically either forward to the next site $k + 1$ with rate $\omega_f_0$, or backward to site $k - 1$ with rate $\omega_b$. Otherwise, if the site $k - 1$ is occupied with a chaperone, the wall will have no choice but to hop forward stochastically by one step and the corresponding rate is denoted as $\omega_f_1$. In summary, the stochastic motion of a membrane along a polymer chain can be divided into two categories, one is the usual diffusion process with possible forward and backward hopping, the other is the rectified diffusion with possible forward hopping alone, since the backward hopping is blocked by chaperones. In this sense, our model can be regarded as a Brownian ratchet translocation model.

We assume that chaperones can stochastically bind to any unoccupied site behind the wall with rate $\omega_a$. At the same time, any attached chaperone can detach from its binding sites at random as well, where the detachment rate is denoted as $\omega_d$. It is easy to find that the dynamics of a translocation process depend on the states of all the binding sites of the polymer chain behind the wall. An ideal model, therefore, should include the states of all these binding sites, i.e. whether it is occupied with some chaperone or not. However, keeping all these states explicitly in the model will make further
theoretical analysis inaccessible, especially for long polymer chains, since the state space grows exponentially with the number of binding sites.

Taking this into consideration, we assume that the binding sites of two units or more away from the wall have sufficient time to reach equilibrium, and the probability of staying unoccupied thereby is \( q = \omega_d/(\omega_a + \omega_d) \). Actually, the binding/unbinding kinetics of chaperones are much faster than the forward/backward diffusion process of a polymer chain’s translocation across the membrane. The ratio of their timescales lies around \( 1/300 \) [36], which therefore makes the assumption plausible.

Our model, characterizing the stochastic translocation of a polymer chain across membrane, or equivalently the stochastic motion of a wall along one-dimensional lattice of length \( N \) is depicted in figure 1. For \( k = 1, \ldots, N \), we denote \( P_{k,0} \) and \( P_{k,1} \) as the probability that the wall stands at site \( k \) with the previous site \( k - 1 \) unoccupied or occupied respectively, see figure 1(a) and (b). The first site of a polymer chain is labeled as 0, and the corresponding probability that the wall stands at this site, site 0, is denoted as \( P_0 \). Additionally, we denote \( Q \) as the probability that none of the sites of a polymer chain is passing through the pore, i.e. the nanopore of the membrane is vacant. In other words, \( Q \) is the probability that a polymer chain has just accomplished its translocation, and a new translocation process has not started yet.

Since chaperones only exist in the target region, only the sites behind the wall are permitted to be bound. Then, the wall, currently standing at site \( k \), can always move forward stochastically to the next site \( k + 1 \), while it gets quite different for backward motions. If the site \( k - 1 \) is unoccupied, the wall at site \( k \) can also stochastically move backward to site \( k - 1 \) with rate \( \omega_b \). However, if site \( k - 1 \) is occupied with a chaperone, the backward motion of the wall at site \( k \) will be blocked. That is to say, the forward motion of the wall standing at site \( k \) may depend on the state of site \( k - 1 \). To state a general case, the forward hopping rate of the wall is denoted differently, where \( \omega_{f_0} \) and \( \omega_{f_1} \) match the cases that site \( k - 1 \) stays in an unoccupied and occupied state.

To picture the site-by-site translocation process, we can still imagine a wall standing at some site on the main body, labeled as \( k \). As described in the beginning of this section, the probability that site \( k - 2 \) stays in an unoccupied state is \( q \), with this site reaching the equilibrium of binding/unbinding kinetics. Based on a sustained translocation process, we establish a one-dimensional system of period \( l \), where one period indicates a complete transfer of one identical polymer chain. Then, master equations of probabilities \( P_{k,0} \) and \( P_{k,1} \) in the main body of a lattice, i.e. \( k = 2, \ldots, N - 1 \), are given by

\[
\frac{\partial P_{k,0}(l, t)}{\partial t} = \omega_{f_0} P_{k-1,0}(l, t) + \omega_{f_1} P_{k-1,1}(l, t) + \omega_d P_{k,1}(l, t) + q \omega_b P_{k+1,0}(l, t) - (\omega_b + \omega_a + \omega_{f_0}) P_{k,0}(l, t),
\]

(1)

\[
\frac{\partial P_{k,1}(l, t)}{\partial t} = \omega_a P_{k,0}(l, t) + (1 - q) \omega_b P_{k+1,0}(l, t) - (\omega_d + \omega_{f_1}) P_{k,1}(l, t).
\]

(2)

See figure 1(c) for the schematic depiction.

Since site 0 is the starting point of translocation process, see figure 1(d), there is no lattice site behind the wall if it stands at site 0, indicating that no chaperone can
be attached to the polymer chain when \( k = 0 \). Thus, the forward hopping rate of the wall at site 0 may be different from both \( \omega_f_0 \) and \( \omega_f_1 \). A different notation \( \omega_f \) is used to denote the forward hopping rate of the wall located at site 0. As schematically depicted in figure 1(d), equation (2) still holds for \( k = 1 \) while equation (1) should be modified as

\[
\frac{\partial P_{1,0}(l, t)}{\partial t} = \omega_f P_{0}(l, t) + \omega_d P_{1,1}(l, t) + q\omega_b P_{2,0}(l, t) - (\omega_b + \omega_a + \omega_f_0) P_{1,0}(l, t).
\]

When it comes to the boundary sites, things change. As discussed above, we use \( Q \) to denote the probability that the system stays in the interval between two translocation processes of separate polymer chains, and in this state, we can imagine the wall is standing at a fictitious site \( Q \). Translocation starts with rate \( \omega_s \) when the wall binds to the initial site 0 of a polymer chain, or we say the wall jumps from the fictitious site \( Q \) to site 0 with rate \( \omega_s \). Similarly, it is when the wall hops forward from the terminal site \( N \) to the fictitious site with rate \( \omega_s \) that the translocation of a polymer chain comes to an end. See figure 1(d) for a schematic depiction. Based on this scenario, the master equation for probability \( P_0 \) is as follows,

\[
\frac{\partial P_0(l, t)}{\partial t} = \omega_s Q(l, t) + \omega_b P_{1,0}(l, t) - \omega_f P_0(l, t).
\]

Similarly, probabilities \( P_{N,0} \) and \( P_{N,1} \) are governed by

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Mean velocity and effective diffusion constant for translocation of biopolymer chains across membrane

\[ \frac{\partial P_{N,0}(l,t)}{\partial t} = \omega_f P_{N-1,0}(l,t) + \omega_f P_{N-1,1}(l,t) + \omega_d P_{N,1}(l,t) - (\omega_b + \omega_a + \omega_e) P_{N,0}(l,t), \]  

\[ \frac{\partial P_{N,1}(l,t)}{\partial t} = \omega_d P_{N,0}(l,t) - (\omega_d + \omega_e) P_{N,1}(l,t). \]  

Once a polymer chain has been transferred to the target region completely, the translocation process enters into a pause state, where the wall will wait until it walks onto another polymer chain, that is when another polymer chain starts its translocation. With figure 1(d), the probability \( Q \) satisfies

\[ \frac{\partial Q(l,t)}{\partial t} = \omega_e [P_{N,0}(l-1,t) + P_{N,1}(l-1,t)] - \omega_s Q(l,t). \]  

Hence, we have setup a system of \( 2N + 2 \) states, where the translocation of a polymer chain across a nanopore of membrane is mapped into the motion of a wall through a one-dimensional lattice in terms of relative motion.

3. Expressions for mean velocity and effective diffusion constant

In this section, we will generalize the main idea of Derrida [35, 37] to realize analytical expressions for mean velocity and effective diffusion constant of translocation of polymer chains across a membrane in the stationary-state limit, \( t \to \infty \). We begin with a general expression for the mean translocation of the wall,

\[ \langle x(t) \rangle = \sum_{l=-\infty}^{+\infty} (NL)Q(l,t) + \sum_{k=0}^{N} (k + NL)P_k(l,t). \]  

That is to say, if we assume the biopolymer chain is of length \( N \), each translocation period will add \( N \) units to the accumulated distance that the wall travels, since hopping away from a polymer chain (translocation termination, from site \( N \) to pause state, or stepping into a new period (translocation initiation, from fictitious site \( Q \) to site 1), does not increase the total distance that the wall travels.

Note that, in equation (8) \( P_k(l,t) := P_{k,0}(l,t) + P_{k,1}(l,t) \) for \( 1 \leq k \leq N \), is the probability that the wall stands at site \( k \) of the \( l \)-th polymer at time \( t \), see figure 1. We now define a few auxiliary functions for each state concerned, they are

\[ B_Q(t) = \sum_{l=-\infty}^{+\infty} Q(l,t), \quad C_Q(t) = \sum_{l=-\infty}^{+\infty} (NL)Q(l,t), \]

\[ B_k(t) = \sum_{l=-\infty}^{+\infty} P_{k,0}(l,t), \quad C_k(t) = \sum_{l=-\infty}^{+\infty} (k + NL)P_{k,0}(l,t), \]  

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Mean velocity and effective diffusion constant for translocation of biopolymer chains across membrane

\[ \tilde{B}_k(t) = \sum_{l=-\infty}^{+\infty} P_{k,1}(l,t), \quad \tilde{C}_k(t) = \sum_{l=-\infty}^{+\infty} (k + N)P_{k,1}(l,t). \]  

Notably, functions in equation (9) are defined for index \( k \) such that \( 0 \leq k \leq N \) whereas those in equation (10) are defined for \( 1 \leq k \leq N \) only. Here, for convenience, we assume \( P_{k,0}(l,t) = P_k(l,t) \) for \( k = 0 \) in equation (9). Generalizing Derrida’s original method [35, 37], we propose the following ansatz in the stationary-state limit, i.e.

\[ B_Q(t) \rightarrow b_Q, \quad B_k(t) \rightarrow \tilde{b}_k, \quad \tilde{B}_k(t) \rightarrow \tilde{\tilde{b}}_k, \]

\[ C_Q(t) \rightarrow a_Q t + T_Q, \quad C_k(t) \rightarrow a_k t + T_k, \quad \tilde{C}_k(t) \rightarrow \tilde{a}_k t + \tilde{T}_k. \]

Obviously, all these \( b_k \)'s together with \( \tilde{b}_k \)'s are governed by normalization condition

\[ b_Q + b_0 + \sum_{k=1}^{N} (b_k + \tilde{b}_k) = 1, \]

since \( b_k \) (or \( \tilde{b}_k \)) gives the probability of finding the wall at a specific state at large time.

In the stationary-state limit, (11), that is \( dB_k/dt = 0 \) and \( d\tilde{B}_k/dt = 0 \) when time \( t \) is infinite, master equations given in section 2 are transformed into

\[ 0 = \omega_s (b_N + \tilde{b}_N) - \omega_s b_Q, \]

\[ 0 = \omega_s b_Q + \omega_d b_1 - \omega_f b_0, \]

\[ 0 = \omega_f b_0 + \omega_d \tilde{b}_1 + q \omega_d b_2 - (\omega_b + \omega_f + \omega_a) b_1, \]

\[ 0 = \omega_f b_1 + \omega_d \tilde{b}_1 + \omega_d \tilde{b}_k + q \omega_d \tilde{b}_k+1 - (\omega_a + \omega_f + \omega_b) b_k, \quad (k = 2, \ldots, N - 1), \]

\[ 0 = \omega_f b_{N-1} + \omega_d \tilde{b}_{N-1} + \omega_d \tilde{b}_N - (\omega_a + \omega_c + \omega_b) b_N, \]

\[ 0 = \omega_c b_k + (1 - q) \omega_c b_{k+1} - (\omega_d + \omega_f) \tilde{b}_k, \quad (k = 1, \ldots, N - 1), \]

\[ 0 = \omega_c b_N - (\omega_d + \omega_c) \tilde{b}_N. \]

These equations lead to a recurrence

\[ Rb_{k+1} - Sb_k = \omega_b b_1 - \omega_f b_0 = -\omega_s b_Q, \quad \text{for} \quad k = 1, \ldots, N - 1, \]

where

\[ R = \omega_b \frac{\omega_d + q \omega_f}{\omega_d + \omega_f}, \quad S = \omega_f \frac{\omega_d + q \omega_f}{\omega_d + \omega_f}. \]

Equation (15) means that the probability-fluxes between each two neighboring effective sites remain constant in steady state.

Considering the stationary state for boundary sites, i.e. the first equation and the last equation in equation (14), we have

\[ b_N = \left[ \frac{\omega_c}{\omega_c + \frac{\omega_s \omega_c}{\omega_d + \omega_c}} \right]^{-1} \omega_c b_0 \equiv K \omega_s b_Q. \]
Mean velocity and effective diffusion constant for translocation of biopolymer chains across membrane

The recurrence given in equation (15) along with equation (17) yields a general expression for \( b_k \),

\[
b_k = \begin{cases} \[ \tilde{S}^{k-N}[K - (S - R)^{-1}] + (S - R)^{-1} \right] \omega_s b_Q, & (k = 1, \ldots, N), \\ \omega_f^{-1}(\omega_f b_1 + \omega_s b_Q), & (k = 0), \end{cases}
\]

(18)

where \( \tilde{S} = S/R \).

Thus, all of \( b_k \)'s and \( \tilde{b}_k \)'s are determined by \( \omega_s b_Q \), which is the probability flux exactly. With the normalizing condition, equation (13), we obtain

\[
\omega_s b_Q = \left[ W_1 + W_2 \tilde{S}^{1-N} + W_3 N \right]^{-1},
\]

(19)

where

\[
W_1 = \frac{1}{\omega_s} + \frac{1}{\omega_f} + \frac{U}{S - R} + K \left( \frac{\omega_a}{\omega_d + \omega_e} - \frac{\omega_a}{\omega_d + \omega_f} \right) + M \frac{S}{S - R} (K - \frac{1}{S - R}),
\]

\[
W_2 = \left( K - \frac{1}{S - R} \right) \left( U - \frac{M}{S - 1} \right) \quad \text{and} \quad W_3 = \frac{M}{S - R},
\]

with

\[
M = 1 + \frac{\omega_a}{\omega_d + \omega_f} + \frac{(1 - q)\omega_b}{\omega_d + \omega_f}, \quad U = \frac{\omega_b}{\omega_f} - \frac{(1 - q)\omega_b}{\omega_d + \omega_f}.
\]

So far, \( b_Q \) has been obtained explicitly as a function of all constant rates. So do \( b_k \)'s and \( \tilde{b}_k \)'s.

Let us return to the ansatz (12) to derive explicit expressions for \( a_k \), \( \tilde{a}_k \), \( T_k \) and \( \tilde{T}_k \). With the definitions of \( C(t) \) and \( \tilde{C}(t) \), i.e. (9) and (10), as well as the master equations presented in section 2, it is easy to show that \( C(t) \) and \( \tilde{C}(t) \) satisfy

\[
\frac{dC_k}{dt}(t) = \omega_e (C_N + \tilde{C}_N) - \omega_s C_Q,
\]

\[
\frac{dC_0}{dt}(t) = \omega_s C_Q + \omega_f C_1 - \omega_f C_0 - \omega_b B_1,
\]

\[
\frac{dC_1}{dt}(t) = \omega_f C_0 + \omega_d \tilde{C}_1 + q \omega_b C_2 - (\omega_b + \omega_a + \omega_f) C_1 + \omega_f B_0 - q \omega_b B_2,
\]

\[
\frac{dC_k}{dt}(t) = \omega_j C_{k-1} + \omega_f \tilde{C}_{k-1} + \omega_d \tilde{C}_k + q \omega_b C_{k+1} - (\omega_b + \omega_a + \omega_f) C_k
\]

\[
+ \omega_f B_{k-1} + \omega_f \tilde{B}_{k-1} - q \omega_b B_{k+1}, \quad (k = 2, \ldots, N - 1),
\]

\[
\frac{dC_N}{dt}(t) = \omega_j C_{N-1} + \omega_f \tilde{C}_{N-1} + \omega_d \tilde{C}_N - (\omega_b + \omega_a + \omega_e) C_N
\]

\[
+ \omega_f B_{N-1} + \omega_f \tilde{B}_{N-1},
\]

\[
\frac{d\tilde{C}_k}{dt}(t) = \omega_a C_k + (1 - q) \omega_b C_{k+1} - (\omega_d + \omega_f) \tilde{C}_k
\]

\[
- (1 - q) \omega_b B_{k+1}, \quad (k = 1, \ldots, N - 1),
\]

\[
\frac{d\tilde{C}_N}{dt}(t) = \omega_a C_N - (\omega_d + \omega_e) \tilde{C}_N.
\]
Mean velocity and effective diffusion constant for translocation of biopolymer chains across membrane

Let time $t \rightarrow \infty$ and focus on the terms proportional to time $t$. Then we get a set of equations analogous to equation (14). Specifically, these equations on $a_k s(\tilde{a}_k s)$ are just the same as equation (14), if $b_k s(\tilde{b}_k s)$ in equation (14) are all replaced by $a_k s(\tilde{a}_k s)$. Thus, one can immediately conclude that

$$a_Q = Ab_Q, \quad a_k = Ab_k, \quad \tilde{a}_k = \tilde{A}b_k,$$

and

$$a_Q + a_0 + \sum_{k=1}^{N}(a_k + \tilde{a}_k) = A[b_Q + b_0 + \sum_{k=1}^{N}(b_k + \tilde{b}_k)] = A,$$  (22)

where $A$ is a constant. Sum up all of the equations in the stationary-state limit of equation (20) and we have

$$A = \omega_f b_0 + \omega_f^0 \sum_{k=1} b_k + \omega_f \sum_{k=1} \tilde{b}_k - \omega_b \sum_{k=1} b_k$$

$$= \omega_b b_0 - \omega_f b_1 + \sum_{k=1}^{N-1}(Sb_k - Rb_{k+1})$$

$$= N\omega_s b_0,$$  (23)

where the last equation follows the recurrence in equation (15) and $b_Q$ is given in equation (19). Therefore, constant $A$ is determined by all of the constant transition rates. Explicit expressions for $a_k s$ and $\tilde{a}_k s$ can also be obtained via equations (18), (19), (21), (23).

Next, we begin to derive the expressions for $T_k s$ and $\tilde{T}_k s$. In the stationary-state limit, equation (20) employs the following expressions,

$$a_Q = \omega_c(T_N + \tilde{T}_N) - \omega_s T_Q,$$

$$a_0 = \omega_s T_Q + \omega_b T_1 - \omega_f T_0 - \omega_b b_1,$$

$$a_1 = \omega_f T_0 + \omega_d \tilde{T}_1 + q\omega_b T_2 - (\omega_b + \omega_s + \omega_f_0)T_1 + \omega_f b_0 - q\omega_b b_2,$$

$$a_k = \omega_f^0 T_{k-1} + \omega_f \tilde{T}_{k-1} + \omega_d \tilde{b}_k + q\omega_b T_{k+1} - (\omega_b + \omega_s + \omega_f_0)T_k$$

$$+ \omega_f b_{k-1} + \omega_f \tilde{b}_{k-1} - q\omega_b b_{k+1}, \quad (k = 2, \ldots, N - 1),$$

$$a_N = \omega_f^0 T_{N-1} + \omega_f \tilde{T}_{N-1} + \omega_d \tilde{T}_N - (\omega_b + \omega_s + \omega_e)T_N$$

$$+ \omega_f b_{N-1} + \omega_f \tilde{b}_{N-1},$$

$$\tilde{a}_k = \omega_a T_k + (1 - q)\omega_b T_{k+1} - (\omega_d + \omega_f_1)\tilde{T}_k - (1 - q)\omega_b b_{k+1},$$

$$\tilde{a}_N = \omega_a T_N - (\omega_d + \omega_e)\tilde{T}_N.$$  (24)

Here, in the stationary-state limit, attention is paid to the terms independent of time $t$ and thus equation (20) reduces to (24).
With some new auxiliary quantities
\[ y_k \equiv \begin{cases} 
\omega_f T_0 - \omega_b T_1, & \text{for } k = 0 \\
ST_k - RT_{k+1}, & \text{for } k = 1, \ldots, N - 1.
\end{cases} \]

Equation (24) can be transformed into a recurrence, that is
\[ y_k - y_{k-1} = Z_k, \quad (25) \]
and the first term comes as
\[ y_0 = \omega_s T_Q + Z_0. \quad (26) \]

Since we have already obtained the explicit expressions for \( b_k \)s as well as \( a_k \)s, \( Z_k \)s turn to known terms, see equation (A.1).

Through a simple summation, it produces
\[ y_k = \omega_s T_Q + r_k, \quad (27) \]
where
\[ r_k = \sum_{i=0}^{k} Z_i, \]
and \( k = 0, \ldots, N - 1 \). See equation (A.6) for expressions of \( r_k \).

It means that each \( y_k \) consists of two components: one has been determined explicitly and the other depends on the undetermined constant \( T_Q \). In fact, the same holds for each \( T_k \), which can be shown through backward iterations. That is
\[ T_k = \begin{cases} 
K(a_Q + \omega_s T_Q) + (\omega_a + \omega_d + \omega_e)^{-1}a_N, & \text{for } k = N, \\
S^{-1}(RT_{k+1} + y_k), & \text{for } k = 1, \ldots, N - 1, \\
\omega_f^{-1}(\omega_b T_1 + y_0), & \text{for } k = 0.
\end{cases} \quad (28) \]

Obviously, \( T_N \) and consequently all \( T_k \)s are made up of these two components as well, i.e. one has been determined explicitly by constant transition rates and the other depends on the undetermined constant \( T_Q \).

We can now write \( T_k = X_k + Y_k \), where \( X_k \) is proportional to \( T_Q \) and \( Y_k \) is described in some determined factors. Particularly, \( T_Q = X_Q \) and \( Y_Q = 0 \). At the same time, \( \tilde{T}_k \)s can be derived from the last two equations in equation (24) and they are shown as
\[ \tilde{T}_k = \begin{cases} 
(\omega_d + \omega_e)^{-1}[\omega_a T_k - \tilde{a}_k], & \text{for } k = N, \\
(\omega_b + \omega_f)^{-1}[\omega_a T_k + (1 - q)\omega_f T_{k+1} - \tilde{a}_k - (1 - q)b_{k+1}], & \text{for } k = 1, \ldots, N - 1.
\end{cases} \quad (29) \]

That is to say, each \( \tilde{T}_k \) can also be split into those two parts and we can then write \( \tilde{T}_k = \tilde{X}_k + \tilde{Y}_k \) with \( \tilde{X}_k \) proportional to \( T_Q \) and \( \tilde{Y}_k \) determined.

Surprisingly, the undetermined constant \( T_Q \) will cancel out in the final expressions for the dynamic properties concerned. Keeping this in mind, we first let \( T_Q = 0 \) for the calculation of \( Y_k \) and show how that works later.

With \( T_Q = 0 \), that is when both \( X_k \) and \( \tilde{X}_k \) equal zero, we find
Mean velocity and effective diffusion constant for translocation of biopolymer chains across membrane

\[ Y_N = K a_Q + (\omega_a + \omega_d + \omega_e)^{-1} \bar{a}_N, \]

\[ Y_k = \tilde{S}^{k-N} Y_N + S^{-1} \sum_{i=0}^{N-1-k} \frac{r_{i+k}}{S^i} \text{ for } k = 1, \cdots, N - 1, \tag{30} \]

\[ Y_0 = \omega_f^{-1} [\omega_b Y_1 + Z_0], \]

\[ Y_Q = 0, \]

and \( \bar{Y}_k \) is given by \( Y_k \) in the same form as equation (29), i.e.

\[ \bar{Y}_k = \begin{cases} (\omega_d + \omega_e)^{-1} [\omega_a Y_k - \bar{a}_k], & \text{for } k = N, \\ (\omega_d + \omega_f)^{-1} [\omega_a Y_k + (1 - q)\omega_f Y_{k+1} - \bar{a}_k - (1-q)b_{k+1}], & \text{for } k = 1, \cdots, N - 1. \end{cases} \tag{31} \]

Now, we are fully prepared to derive analytical expressions for dynamic properties of interest, i.e. the mean translocation velocity \( V \) and effective diffusion constant \( D \) in the steady state, which is respectively defined as

\[ V = \lim_{t \to +\infty} \frac{d \langle x(t) \rangle}{dt}, \tag{32} \]

and

\[ D = \lim_{t \to +\infty} \frac{1}{2} \frac{d}{dt} [\langle x^2(t) \rangle - \langle x(t) \rangle^2]. \tag{33} \]

With equation (8) and the auxiliary functions given in equations (9) and (10),

\[ \frac{d \langle x(t) \rangle}{dt} = \frac{d C_0(t)}{dt} + \sum_{k=0}^{N} \frac{d C_k(t)}{dt} + \sum_{k=1}^{N} \frac{d \tilde{C}_k(t)}{dt}, \tag{34} \]

which is exactly the summation of equation (20). Recall the derivation of constant \( A \) (see equations (21)–(23)), as well as the stationary-state assumption as is given in equation (12), and we can easily get the mean velocity in the stationary-state limit, i.e.

\[ V = a_Q + \sum_{k=0}^{N} a_k + \sum_{k=1}^{N} \bar{a}_k = A = N \omega_s b_Q, \tag{35} \]

where the probability flux \( \omega_s b_0 \) is given by (19).

An explicit expression for effective diffusion constant \( D \) requires more detailed analysis. Utilizing the master equations in section 2, the differentiation of \( \langle x^2(t) \rangle \) turns to

\[ \frac{1}{2} \frac{d \langle x^2(t) \rangle}{dt} = D_1 + D_2, \tag{36} \]

where

\[ D_1 = \left[ \omega_f C_0 + \omega_f b_0 \sum_{k=1}^{N-1} C_k + \omega_f b_1 \sum_{k=1}^{N-1} \bar{C}_k - \omega_b \sum_{k=1}^{N} C_k \right], \tag{37} \]
\[ D_2 = \frac{1}{2} \left[ \omega_f B_0 + \omega_{f_0} \sum_{k=1}^{N-1} B_k + \omega_{f_1} \sum_{k=1}^{N-1} \tilde{B}_k + \omega_b \sum_{k=1}^{N} B_k \right]. \]  

(38)

At the same time, the second part of \( D \) as given in (33) can be described as

\[ D_3 = \frac{1}{2} \frac{d(x(t))^2}{dt} = V \langle x(t) \rangle = A \left[ C_Q + \sum_{k=0}^{N} C_k + \sum_{k=1}^{N} \tilde{C}_k \right]. \]  

(39)

In the stationary-state limit, we first claim that terms proportional to time \( t \) in the expression for \( D \) cancel out. Since \( B_k(\tilde{B}_k) \) tends to be a constant \( b_k(\tilde{b}_k) \) as time \( t \) goes infinity, we just concentrate on equations (37) and (39) to verify it. Replacing \( C_k(\tilde{C}_k) \) with \( a_k(\tilde{a}_k) \) in equation (37), one can easily find that the coefficient of \( t \) in \( D_1 \) comes as \( A \cdot N\omega_s b_Q = A^2 \), as compared with equation (23). Similarly, the coefficient of \( t \) in \( D_3 \) is

\[ A \left[ a_Q + \sum_{k=0}^{N} a_k + \sum_{k=1}^{N} \tilde{a}_k \right] = A^2. \]

Then, we focus on the terms proportional to \( T_Q \) in order to show that the effective diffusion constant \( D \) is independent of them. As mentioned before, each \( T_k \) and \( \tilde{T}_k \) can be written as \( T_k = X_k + Y_k \) and \( \tilde{T}_k = \tilde{X}_k + \tilde{Y}_k \), respectively. With a glance at equations (27) and (28), we find \( X_k \) follows the same recurrence as \( b_k \) does. In other words, every \( b_k \) equation (15) can be replaced with \( X_k \). So does \( \tilde{X}_k \) with respect to \( \tilde{b}_k \). Thus, we have

\[ X_k = \frac{T_{Q}}{b_Q} b_k, \quad \tilde{X}_k = \frac{T_{Q}}{b_Q} \tilde{b}_k. \]  

(40)

This can be better understood if the procedure of seeking \( b_k \) and \( \tilde{b}_k \) is regarded as solving linear equations \( \Omega x = 0 \) corresponding to equation (14), where \( \Omega \) is a matrix of \( 2(N + 1) \times 2(N + 1) \) in size. Since the rank of \( \Omega \) is \( 2N + 1 \), \( b = (b_Q, b_0, b_1, b_2, \cdots, b_N, \tilde{b}_N)^\text{T} \), what we obtained in the foregoing is determined on the normalizing condition. If we denote \( T \) as the vector \((T_Q, T_0, T_1, \tilde{T}_1, \cdots, T_N, \tilde{T}_N)^\text{T}\), one can find the derivation of \( T \) is just solving the linear equation \( \Omega x = \beta \) due to equation (24), where each component in \( \beta \) is determined by \( b_k(\tilde{b}_k) \) and \( a_k(\tilde{a}_k) \). Actually, we have already obtained a solution for this equation, which is \( Y = (Y_Q, Y_0, Y_1, \tilde{Y}_1, \cdots, Y_N, \tilde{Y}_N)^\text{T} \) with each component given in equations (30) and (31).

Then, a general solution \( T \) can be written as \( T = \alpha b + \tilde{Y} \). Particularly, the first component in \( Y \) is zero, i.e. \( Y_Q = 0 \). If we regard \( T_Q \) as a undetermined constant, the undetermined constant should be restricted to \( \alpha = T_Q/b_Q \) in the consideration of the first component in \( T, Y \) and \( b \). Equation (40) holds therefore. That is exactly what we have done.

In this way, considering whether the terms proportional to \( T_Q \) in equations (37) and (39) can cancel out is to examine exactly the performance of \( X_k \). It is easy to see such terms in the former equation sum up to \( N\omega_s T_Q \), if we take \( X_k \) as a substitution for \( C_k \) in equation (37) and compare the coefficients with those in equation (23). The latter
Mean velocity and effective diffusion constant for translocation of biopolymer chains across membrane

equation (39), leads to a summation of $X_k$’s and $\tilde{X}_k$’s, which is $N\omega_s T_Q$ as well. As a result, terms proportional to $T_Q$ cancel out.

Due to these two cancellations, we finally derive the explicit expression for effective diffusion constant in the stationary-state limit,

$$D = D_1 + D_2 - D_3 = -\sum_{k=0}^{N-1} s_k - \sum_{k=1}^{N-1} \tilde{s}_k + \frac{V}{2} N - V \left[ Y_0 + \sum_{k=1}^{N} (Y_k + \tilde{Y}_k) \right],$$

where $Y_k$ has already been obtained in equation (30), $\tilde{Y}_k$ follows equation (31) and

$$s_k = \sum_{i=1}^{k} a_i, \quad \tilde{s}_k = \sum_{i=2}^{k} \tilde{a}_i.$$

See appendix A for more details.

4. Properties of polymer translocation across membrane

To further understand the analytical results and get an intuition about the translocation process of polymer chains across a membrane, extensive numerical explorations are carried out according to explicit results given in equations (35) and (41). Instead of general analysis with respect to each transition rate, particular attention is paid to a special case where $\omega_f = \omega_f = \omega_b = \omega_b \equiv w$, due to the fact that the forward/backward diffusion of a polymer chain essentially comes from Brownian fluctuations. This means that it is the binding of chaperones that rectifies the diffusion of polymer chain to a directional motion on average. In fact, results remain valid even if the polymer chain undergoes a biased random walk when crossing the membrane. Note that both the binding and unbinding of chaperones are extremely fast compared to the forward/backward diffusion of a polymer chain. In all calculations below, we always keep the binding/unbinding rate of a chaperone at least 2 orders of magnitude higher than the forward/backward diffusion rate.

Numerical results on mean translocation velocity $V$ are displayed in figure 2. When trailing binding sites (i.e. binding sites of a polymer chain lying in the target region) are occupied with chaperones in large probability, the polymer chain with a larger forward/backward diffusion rate will pass through the pore more rapidly, since its backward motion is hindered by chaperones. So translocation velocity increases with the diffusion rate $w$. This rise, however, will not last indefinitely, since for sufficiently large diffusion rate $w$, the translocation velocity will be limited by the binding/unbinding process of chaperones as well as the finite starting rate $\omega_s$ and ending rate $\omega_e$. See figure 2(a).

It means that there is an upper limit for mean velocity $V$ as diffusion rate $w$ tends to infinity, see figure 2(a). Besides, we notice that a longer polymer chain employs greater mean velocity, provided large diffusion rate $w$ and high ratio $\omega_a/\omega_d$. This matches the fact that the pretty slow initiation process and termination process, acting as restrictions, produce greater impact on the overall mean velocity of a shorter polymer chain.

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Mean velocity and effective diffusion constant for translocation of biopolymer chains across membrane

From another perspective, we consider the mean dwell time $t$ for some translocation, which is

$$t = \frac{N}{V} = W_1 + W_2 S^{1-N} + W_3 N. \quad (42)$$

Here, the numerator $N$ is the length of a polymer chain and the denominator $V$ is the mean velocity of translocation. It reflects the effect of binding sites’ number on polymers of fixed length. According to the definition of constant $S$, if the diffusion rate $w$ is low, $S$ increases almost linearly with $w$, while $W_1, W_2, W_3$ are all insensitive to the low diffusion rate $w$. Therefore, equation (42) indicates that the relation between dwell time $t$ and diffusion rate $w$ follows power law approximately, when the polymer chain diffuses at a low diffusion rate $w$. See figure 2(b).

The dependence of mean velocity $V$ on the ratio of binding/unbinding rate of a chaperone is presented in figure 2(c). With high ratio $\omega_a/\omega_d$, the binding site lying in the target region will be more likely to be occupied with a chaperone, which will, then, help rectify the forward/backward diffusion. That is, high binding/unbinding ratio speeds up the translocation of a polymer chain. The dependence of mean velocity on ratio $\omega_a/\omega_d$ displays similar performance to that on diffusion rate $w$, as is shown in figures 2(a) and (c). If the ratio $\omega_a/\omega_d$ is large enough, mean translocation velocity $V$ increases with the length $N$ of a polymer chain and vice versa. See figure 2(c) and the inset. This is because the longer a polymer chain is, the less influence is likely to be exerted through the relatively low starting rate $\omega_s$ and ending rate $\omega_e$. Conversely, low ratio $\omega_a/\omega_d$ leads to the slow translocation of the main body part of a polymer chain, and the relatively large starting rate $\omega_s$ as well as ending rate $\omega_e$ plays a more active role in promoting the overall mean translocation velocity $V$ of a shorter polymer chain.

It means that the mean velocity will decrease with the growth of the polymer length $N$.

Note that, experimental binding/unbinding ratio is approximate to 833.3 [27]. Figure 2(c) shows that the velocity $V$ is almost independent of ratio $\omega_a/\omega_d$ when $\omega_a/\omega_d \geq 833.3$, implying that the translocation process is limited by the forward/backward diffusion rate $w$ as well as starting rate $\omega_s$ and ending rate $\omega_e$. Moreover, the plots in figures 2(a) and (b) show that, for diffusion rate $w$ larger than 1000 s$^{-1}$, mean velocity $V$ is also insensitive to the change of rate $w$. Therefore, the translocation of polymer chain is limited merely by the starting and the ending process when $\omega_a/\omega_d \geq 833.3$ and diffusion rate $w \geq 1000$ s$^{-1}$. Obviously, velocity $V$ increases with both rate $\omega_s$ and rate $\omega_e$, see figure 1(d) or equations (19), (35). Note that, in figures 2(a) and (b), the ratio $\omega_a/\omega_d$ is always kept 833.3.

The dependence of dwell time $t$ on length $N$ of a polymer chain, with different values of ratio $\omega_a/\omega_d$, is plotted in figure 2(d). One can easily find that $t$ increases almost linearly with $N$, especially for large binding/unbinding ratio $\omega_a/\omega_d$. It can been seen in equation (42) as well. Meanwhile, figure 2(d) also shows that dwell time $t$ decreases with the ratio $\omega_a/\omega_d$, since a polymer chain requires high mean velocity $V$ provided with a great binding/unbinding ratio $\omega_a/\omega_d$. See figure 2(c) and the definition of $t$ given in equation (42).

The discussion above on mean velocity $V$ provides a chief description of the dynamics of polymer translocation across a membrane, but it is not sufficient for many biologically relevant cases, especially when the polymer chain is not long. In these situations,
Effective diffusion constant $D$, or say dispersion, plays an important role. The plots in figure 3(a) show that, for high diffusion rate $w$, diffusion constant $D$ increases with both the ratio $\omega_a/\omega_d$ and the polymer length $N$, when the polymer is not long. At the same time, diffusion constant $D$ tends to its lower limit or upper limit monotonically as the ratio $\omega_a/\omega_d$ tends to 0 or infinity, respectively. However, something interesting happens as diffusion rate $w$ goes down approaching the starting rate $\omega_s$ and ending rate $\omega_e$. The plots in figure 3(b) show that, for such cases, $D$ will no longer change monotonically with rate $\omega_a/\omega_d$. Neither will it increase with the polymer length $N$. With a particular value of ratio $\omega_a/\omega_d = 833.3$ [27], the dependence of $D$ on diffusion rate $w$ is plotted in figure 3(c), where the effective diffusion constant $D$ increases with $w$ as expected. Besides, the saturation level for effective diffusion constant $D$ rises with the polymer length $N$, as long as the polymer chain is not too long.
Compare figures 2(a), (c) with 3(a), (c), and we can find that mean velocity $V$ and effective diffusion constant $D$ show almost the same behavior. Both of them increase significantly when the parameter, either forward/backward diffusion rate $w$ or binding/unbinding ratio $\omega_a/\omega_d$, varies within some range. With the parameter varying beyond this range, however, mean velocity $V$ and effective diffusion constant $D$ both stay almost constant. The reason is that the dynamics of translocation process is regulated jointly by several factors. Besides diffusion rate $w$ along with the ratio $\omega_a/\omega_d$, these factors include starting rate $\omega_s$ and ending rate $\omega_e$. See figure 1. When the corresponding parameter, rate $w$ or $\omega_a/\omega_d$, varies within an appropriate range, the translocation process of a polymer is dominated by its diffusion process and the binding/unbinding process of a chaperone, and the mean velocity or diffusion constant changes significantly with the parameter, rate $w$ or ratio $\omega_a/\omega_d$. Otherwise, the dynamics of translocation process is regulated by several factors. Of these, the diffusion rate along with the ratio $\omega_a/\omega_d$, is most important in determining the mean velocity or diffusion constant. The effective diffusion constant $D$ as a function of the ratio $\omega_a/\omega_d$ is displayed in (a) and (b). The diffusion rate used in (a) is $w = 10^2$ s$^{-1}$, while calculations in (b) are carried out with lower diffusion rates. $w = 1$ s$^{-1}$ in (b) and $w = 10$ s$^{-1}$ in inset. (c) Dependence of $D$ on the diffusion rate $w$, with ratio $\omega_a/\omega_d = 833.3$ as is given in [27]. (d) The randomness, $r = 2D/NV$, as a function of the length $N$ of a polymer chain. Several cases with different values of diffusion rate $w$ and ratio $\omega_a/\omega_d$ are considered. Detailed instructions on parameters are listed in the legend. Similar to figure 2, $\omega_d$ is held of order $10^8$ s$^{-1}$ to make the binding/unbinding process of a chaperone much faster than the forward/backward diffusion process of a polymer chain. Other parameters used in calculations are $\omega_s = 1$ s$^{-1}$ and $\omega_e = 1$ s$^{-1}$.
Mean velocity and effective diffusion constant for translocation of biopolymer chains across membrane

will be limited by rate $\omega_s$ and $\omega_e$, and therefore is almost independent of the rate $w$ and ratio $\omega_a/\omega_d$. In other words, the translocation of a polymer chain across a membrane consists of several kinetics, namely initiation, termination, binding/unbinding of chaperones and forward/backward diffusion of a polymer chain. For a given set of parameters, the overall translocation process may be limited by only one or some of them, with others fast enough to be neglected. But the limit process may be switched from one to another, with the change of one or some parameter values.

It is also noteworthy to compare the degree of fluctuations of this stochastic process. An important dimensionless function to evaluate this quantity is randomness, which is defined as [37, 38]

$$r = \frac{2D/N^2}{V/N} = \frac{2D}{NV}.$$  

The plots in figure 3(d) show that the randomness $r$ always decreases with the polymer length $N$. With the ratio $\omega_a/\omega_d = 833.3$ [27], $r$ increases with the forward/backward diffusion rate $w$. Our results also suggest a decrease of $r$ with the number of binding sites $N$. A rapid decay in randomness $r$ can be observed on the condition of a low diffusion rate $w$, whereas the length of a polymer reduces $r$ slightly, but almost linearly, as long as the diffusion rate $w$ is high.

5. Conclusions and remarks

The Brownian ratchet builds a general framework describing how diffusive motion is rectified by chemical energy. Particularly, the chaperone-assisted translocation of polymer chains across a membrane constitutes a fine example of how directed motion can emerge from random diffusion, where the binding and unbinding of chaperones plays a prominent role.

In this paper, a theoretical model is presented mapping the process of translocation across membrane into the discrete motions of a membrane on the polymer chain, where the probability of finding membrane at some site of polymer chain is governed by usual master equations. With this model, the mean velocity and effective diffusion constant of translocation process can be derived explicitly, allowing us to discuss the dynamics of translocation across membrane much more efficiently.

Based on the exact expressions, detailed discussions on particular cases are presented, where the polymer chain is assumed to diffuse freely around the nanopore embedded in a membrane if there is no chaperone molecule. Our results show that both the increase of forward/backward diffusion rate $w$ of polymer chains and the rise in binding/unbinding rate ratio $\omega_a/\omega_d$ of chaperones raise the mean translocation velocity monotonically to a polymer length dependent limit. With large diffusion rate $w$ or great ratio $\omega_a/\omega_d$, the mean velocity increases with the length of polymer chain, while opposite performance can be observed when $w$ or $\omega_a/\omega_d$ stays at a low level. The effective diffusion constant also increases with the diffusion rate $w$, but it will exhibit complicated properties when regarded as a function of ratio $\omega_a/\omega_d$. Meanwhile, the randomness decreases monotonically with the length of a polymer chain.
For most of biological processes in living cells, mean velocity together with effective diffusion constant is usually sufficient to describe the basic dynamic properties. Nevertheless, higher order moments, or higher cumulants, can be obtained as well, simply via the same methods in this paper. Meanwhile, all of the transition rates are assumed to be independent of the site of a polymer chain in our discussion. But in a promoted model with site-dependent transition rates, explicit expressions for mean velocity and effective diffusion constant can still be obtained sketching the key idea of this study.

Appendix. Derivation of effective diffusion constant equation (41)

Since the final expression for diffusion constant $D$ does not depend on the undetermined constant $T_Q$, we let $T_Q = 0$, which indicates that both $X_i$s and $\tilde{X}_k$s are equal to zero. At the same time, $Y_k$ follows equation (30) and $\tilde{Y}_k$ follows (31).

With equation (24),
$$
Z_0 = -a_0 - \omega b_1,
$$
$$
Z_1 = -a_1 - \frac{\omega_d}{\omega_d + \omega_f} \tilde{a}_1 - R b_2 + \omega_f b_0,
$$
$$
Z_k = -a_k - \frac{\omega_f}{\omega_d + \omega_f} \tilde{a}_{k-1} - \frac{\omega_d}{\omega_d + \omega_f} \tilde{a}_k - R b_{k+1} + S b_{k-1},
$$
\hspace{1cm} (k = 2, \ldots, N - 1).

In the stationary-state limit, equations (37)–(39) turn to
$$
D_1 = \omega_f Y_0 + \omega_f \sum_{k=1}^{N-1} Y_k + \omega_f \sum_{k=1}^{N-1} \tilde{Y}_k - \omega b \sum_{k=1}^{N} Y_k,
$$
\hspace{1cm} (A.2)
$$
D_2 = \frac{1}{2} \left[ \omega_f b_0 + \omega_f \sum_{k=1}^{N-1} b_k + \omega_f \sum_{k=1}^{N-1} \tilde{b}_k + \omega b \sum_{k=1}^{N} b_k \right],
$$
\hspace{1cm} (A.3)
and
$$
-D_3 = -A \left[ Y_0 + \sum_{k=1}^{N} (Y_k + \tilde{Y}_k) \right].
$$
\hspace{1cm} (A.4)

Take equation (29) into equation (A.2),
$$
D_1 = (\omega_f Y_0 - \omega b Y_1) + \sum_{k=1}^{N-1} (S Y_k - R \tilde{Y}_{k+1}) - \frac{\omega_f}{\omega_d + \omega_f} \sum_{k=1}^{N-1} \tilde{a}_k
$$
\hspace{1cm} -(1 - q) \omega b \frac{\omega_f}{\omega_d + \omega_f} \sum_{k=1}^{N-1} b_{k+1},
$$
$$
= -\frac{\omega_f}{\omega_d + \omega_f} \sum_{k=1}^{N-1} \tilde{a}_k - (1 - q) \omega b \frac{\omega_f}{\omega_d + \omega_f} \sum_{k=1}^{N-1} b_{k+1} + \sum_{k=0}^{N-1} r_k.
$$
\hspace{1cm} (A.5)
With the definition of $r_k$, we have
\[
\begin{align*}
 r_0 &= Z_0 = -a_0 - \omega_b b_1, \\
r_1 &= \sum_{i=0}^{1} Z_i = -\sum_{i=0}^{1} a_i - \frac{\omega f_i}{\omega_d + \omega f_1} \tilde{a}_1 - R b_2 + \omega_s b Q, \\
r_k &= \sum_{i=0}^{k} Z_i = -\sum_{i=0}^{k} a_i - \sum_{i=1}^{k-1} \tilde{a}_i - \frac{\omega f_1}{\omega_d + \omega f_1} \tilde{a}_k - R b_{k+1} + k \omega_s b Q, \\
(k = 2, \ldots, N - 1).
\end{align*}
\]

Then,
\[
\begin{align*}
\sum_{k=0}^{N-1} r_k &= -\frac{\omega_d}{\omega_d + \omega f_1} \sum_{k=1}^{N-1} \tilde{a}_k - R \sum_{k=1}^{N} b_k - \omega_b b_1 \\
&\quad + \omega_s b Q \sum_{k=1}^{N-1} k - \sum_{k=0}^{N-1} \sum_{i=0}^{k} a_i - \sum_{k=1}^{N-2} \sum_{i=1}^{k} \tilde{a}_i. \\
&= \omega_b \sum_{k=1}^{N} b_k + \frac{1}{2} N(N-1) \omega_s b Q. \\
\end{align*}
\]

Note that the summation of the first term in equation (A.5) and that in equation (A.7) is
\[
-\frac{\omega f_1}{\omega_d + \omega f_1} \sum_{k=1}^{N-1} \tilde{a}_k - \frac{\omega_d}{\omega_d + \omega f_1} \sum_{k=1}^{N-1} \tilde{a}_k = -\sum_{i=1}^{N-1} \tilde{a}_i,
\]
and the summation of terms related to $b_k$ in these two equations equals
\[
-\omega_b \sum_{k=1}^{N} b_k + \frac{1}{2} N(N-1) \omega_s b Q.
\]

Then
\[
D_1 = -\sum_{k=0}^{N-1} s_k - \sum_{k=1}^{N-1} \tilde{s}_k - \omega_b \sum_{k=1}^{N} b_k + \frac{1}{2} N(N-1) \omega_s b Q, \\
(A.8)
\]
where
\[
s_k = \sum_{i=0}^{k} a_i, \quad \tilde{s}_k = \sum_{i=1}^{k} \tilde{a}_i.
\]

For equation (A.3),
\[
D_2 = \frac{1}{2} \left[ \frac{\omega f b_0 + \omega f_0}{\omega_d + \omega f_1} \sum_{k=1}^{N-1} b_k + \omega f_1 \sum_{k=1}^{N-1} b_k - \omega b \sum_{k=1}^{N} b_k + 2 \omega b \sum_{k=1}^{N} b_k \right] \\
= \omega_b \sum_{k=1}^{N} b_k + \frac{1}{2} V. \\
(A.9)
\]

With (35), the summation of equations (A.8), (A.9) and (A.4) can be written as

https://doi.org/10.1088/1742-5468/ab633d
Mean velocity and effective diffusion constant for translocation of biopolymer chains across membrane

\[
D = D_1 + D_2 - D_3 = - \sum_{k=0}^{N-1} s_k - \sum_{k=1}^{N-1} \tilde{s}_k + N \frac{V}{2} - V \left[ Y_0 + \sum_{k=1}^{N} (Y_k + \tilde{Y}_k) \right],
\]

which is exactly equation (41).

References


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Mean velocity and effective diffusion constant for translocation of biopolymer chains across membrane

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